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Sesame (*Sesamum indicum* **L.) Response to Soil Additives Applied in-Furrow at Planting**

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Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

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Original Research Article

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ABSTRACT

Aims: Studies were conducted to study the response of sesame to soil additives applied in-furrow. **Study Design:** Randomized complete block with 4 replications.

Place and Duration of Study: Field experiments were carried out during the 2016 through 2018 growing seasons in south-central Texas near Yoakum (29.27704° N, -97.12453° W).

Methodology: Sesame seed was planted < 2.54 cm deep. Treatments were applied using a CO₂pressurized sprayer in 46.8 L ha-1 of water with one Teejet® orifice disc #45 nozzle per row immediately after seed drop but prior to furrow closure. Each plot consisted of two rows spaced 97 cm apart and 7.6 m long. Sprinkler irrigation was applied on a 2- to 3-wk schedule throughout the growing season as needed. S-metolachlor at 1.4 kg ha⁻¹ was applied preemergence while clethodim at 0.11 kg ha⁻¹ and diuron at 1.12 kg ha⁻¹ were applied postemergence to control annual grasses and broadleaf weeds that were present.

Results: In 2016, treatments containing 7% N + 10% chelated Fe, gibberellic acid + 3-indolebutyric acid (0.045%) + cytokinin as Kinetin (0.09%), and pop-up fertilizer (9-30-0 + Zn) resulted in the greatest sesame emergence. In 2017, 2% N, bifenthrin + *Bacillus amyloliquefaciens* strain D747,

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and humic acids + *Bacillus* spp. resulted in greater emergence (90-97%) while in 2018, *Azospirillum brasilense* and 2% N resulted in the greater emergence (90-91%). In 2016, 2% N produced the greatest yield while in 2018 2% N and the 3-way combination of cytokinin as kinetin (0.090%) + gibberellic acid + indole-3-butyric acid (0.045%) resulted in up to a 117% increase in yield over the untreated.

Conclusion: The 3-way combination of gibberellic acid + 3-indolebutyric acid (0.045%) + cytokinin as kinetin (0.090%) and 2 % N proved to be the most consistent soil additives and resulted in a yield increases in the two years that the studies were harvested.

Keywords: Soil additives; sesame growers; fungicides; microbial enhancers.

1. INTRODUCTION

"Sesame growers are always attempting to improve their production by making use of new technologies and ideas. Using soil additives which includes fungicides, insecticides, soil activators, soil conditioners wetting agents, inoculants, microbial enhancers, and soil stimulants have been investigated and researched since the beginning of the 20th century" [1,2]. Increases in production costs for growers, especially for fertilizers, has renewed interest in these products.

"The production and application of fertilizers are not only costly but can result in unwanted consequences since they are produced from natural gas and their excessive use causes movement into groundwater, ammonia volatilization, and denitrification" [3]. Soil additives vary from synthetic fertilizers in that they usually do not have any nutrient value and they do not have a quality analysis of their content (e. g., 10-34-0 or 32-0-0) [1,2]. "The literature on such products often suggests that by using soil additives, crop production will increase since root growth and nutrient uptake will be improved and therefore, result in an increase in yield. These improvements to the soil are thought to happen when applications are made at the recommended or near recommended rates; however, some additives also claim to reduce or replace the use for fertilizers" [1,2].

"Soil additives can also be used to improve the texture of the soil. Fertilizers can just add nutrients to the soil; however, some additives can alter the soil condition and also add nutrients. Tilth is the physical condition of the soil and factors which determine tilth include soil texture, structure, fertility, and interaction with the organic content and living soil organisms" [4]. By improved the tilth of the soil, roots can penetrate the soil easier and help with water infiltration [4]. Soil amendments can also change the soil in ways that affect the use of plant nutrients [1,2].

Fertilizers will have an effect on plant growth directly by delivering nutrients to plants while soil amendments or additives can affect growth indirectly. Soil additives can not be considered fertilizer substitutes; however, they can help fertilizers become more effective by improving soil texture and tilth and improving microbial activity. Soil additives are most often separated into three different categories: 1) soil conditioners, 2) soil activators, and 3) wetting agents and surfactants. Soil conditioners are defined as products that can help improve a soil's physical condition or structure which, improves the soil's aeration and water relationships [1,2].

Improving and/or maintaining soil structure is one of the main goals in crop production and adding organic matter is one way to improve soil structure [5]. "Soil additives are sold on the basis that they inoculate the soil with new beneficial organisms or stimulate existing soil microbes [5]. Some manufacturers also suggest that these products may improve the physical properties of soil which includes increasing structure and reducing compaction, increase the uptake of fertilizers and soil nutrients, improve crop yields, improve other soil issues such as salinity, and may also help in disease and insect control/resistance" [6]. "Wetting agents and surfactants have helped reduce the surface tension of spray droplets and increase the coverage of the leaf surface with the application of postemergence (POST)pesticides. Adding surfactants to POST herbicides can be used to reduce the risk of crop injury and also has shown to improve the efficiency of preemergence herbicides that have residual soil activity" [7]. Also, some of these products are marketed on the basis that they will loosen tight or compacted soils, improve water infiltration and retention, enhance nutrient availability, and increase crop yields [8].

Some soil additives have been investigated through research trials to document their benefits and limitations. However, sufficient research funds are frequently not available to study the many new products being marketed. Sesame producers need to be made aware of the products available and have some knowledge of their potential for improved sesame production. Therefore, this study was conducted to evaluate soil additives that are currently on the market to determine sesame growth and yield response.

2. MATERIALS AND METHODS

2.1 Field Studies

These studies were conducted during the 2016 through 2018 growing seasons at the Texas A&M AgriLife Research Site near Yoakum $(29.1642^{\circ}$ N, -97.1243 $^{\circ}$ W) in south-central Texas to evaluate sesame response to soil additives applied in-furrow at planting. The tests were

located in the same general area but different parts of the field in the three test years. Soils were a Denhawken-Elmendorf complex (fine, smectitic, hyperthermic Vertic Ustochrepts) with < 1% organic matter, 25% sand content, 38% clay content, and 37% loam with a pH of 7.8 and a cation exchange capacity (CEC) of 34.

2.2 Soil Additives and Sesame Planting

The soil additives in this study are listed in Table 1. Sesame was planted July 13, 2016, July 5, 2017, and May 9, 2018 using a Monosem® planter calibrated to deliver 320 seed m-1 . The later planting dates in 2016 and 2017 were later due to heavy rains in April, May and early June which prevented timely entry into the field. The sesame variety S-35 was planted in 2016 and 2017 while S-34 was planted in 2018.

Sesame seed was planted < 2.54 cm deep and treatments were applied in-furrow with 46.8 L ha- 1 of water using a CO₂-pressurized sprayer with one Teejet® orifice disc # 45 nozzle per row immediately after seed drop but prior to furrow closure. Each individual plot consisted of two rows spaced 97 cm apart and 7.6 m long. The experimental design was a randomized complete block with four replications. An untreated check was also included in each test. Sprinkler irrigation was applied on a 2- to 3-wk schedule throughout the growing season as needed. *S*metolachlor at 1.4 kg ha⁻¹ was applied preemergence while clethodim at 0.11 kg ha-1 and diuron at 1.12 kg ha⁻¹ were applied postemergence to control annual grasses and broadleaf weeds that were present in the test area. Clethodim was applied prior to sesame bloom to prevent any type of injury to the sesame [9].

2.3 Sesame Stand Counts and Harvest

Sesame emergence or stand was estimated visually on a scale of 0 to 100 ($0 =$ no emergence and 100 = complete emergence) [10]. Emergence was evaluated 7 and 161 days after planting (DAP) in 2016, 5 and 64 DAP in 2017, and 15 DAP in 2018. In 2017, the 64 DAP evaluation was taken after sesame death due to excessive moisture while no late-season evaluation was taken in 2018 due to a limited time schedule. Sesame was harvested at 6% moisture in 2016 (161 DAP) and in 2018 (208 DAP) using an Almaco[®] small-plot combine. Yields were not taken in 2017 due to Hurricane Harvey which came through the area on August 25-29 and dumped over 430 mm of rainfall. This high amount of rainfall killed the sesame.

2.4 Data Analysis

Data for percentage of sesame stand and yield were transformed to the arcsine square root prior to analysis; however, non-transformed means are presented because arcsine transformation did not affect interpretation of the data. Data were subjected to ANOVA and analyzed using the SAS PROC MIXED procedure 23 [11]. Treatment means were separated using Fisher's Protected LSD at $P = 0.05$ and the untreated check was used for all data analysis.

3. RESULTS AND DISCUSSION

3.1 Sesame stand

3.1.1 2016. When evaluated 7 DAP, tebuconazole, *Azospirillum brasilense*, 3indolebutyric acid (0.85%) + cytokinin, as kinetin (0.15%), and 2% N resulted in lower sesame emergence than the untreated check (Table 2). Pop up fertilizer at 46771 ml ha $^{-1}$ resulted in the greatest emergence. At the 161 DAP evaluation, taken just prior to harvest, only 7% N + 10% chelated Fe produced sesame stands greater than the untreated check. Gibberellic acid (0.03%) + 3-indolebutyric acid (0.045%) + cytokinin as kinetin (0.09%) and pop up fertilizer at 46771 ml ha⁻¹ also produced $>$ 90% sesame stands (Table 2). *Azospirillum brasilense* showed a 25% reduction in stand from the untreated check. The lack of a sesame response seen with tebuconazole was surprising because these soils do have a history of seedling diseases [12] and fungicides in furrow at planting has shown to improve seed emergence and early-season vigor in soils with a history of seeding diseases [13]. Phipps [13] also reported in peanut (*Arachis hypogaea* L.) tebuconazole suppressed *Cylindrocladium* black rot (caused by *Cylindrocladium parasiticun*).

However, Jordan et al. [14] found that using tebuconazole in-furrow in peanut showed a slower than normal emergence and a reduction in early-season growth. They found that tebuconazole reduced yield in only one of five experiments even though peanut emergence was delayed in most of the studies and plant diameter was reduced when tebuconazole was applied.

3.1.2 2017.At the 5 DAP evaluation, no sesame had emerged in the untreated check, *Azospirillum brasilense*, 7% N + 10% chelated Fe, ionized sodium silicate family, bifenthrin, popup fertilizer + Zn, pyraclostobin alone, pop-up fertilizer + Zn + pyraclostobin, microalgae, arbuscular mycorrhizal fungi, or humic acids + various strains of *Bacillus* spp. at 3363 gr ha-1 plots (Table 2). Treatments of gibberellic acid (0.03%) + 3-indolebutyric acid (0.45%) + cytokinin as kinetin (0.09%), 2% N, bifenthrin + *Bacillus amyloliquefaciens* strain D747, and humic acids + various strains of *Bacillus* spp. at 1121 gr ha⁻¹ resulted in sesame emergence which ranged from 15 to 35%. At the 64 DAP evaluation, all treatments with the exception of those containing pop-up fertilizer + Zn, resulted in greater stands than the untreated check. Mascagni et al [15] reported in corn (*Zea mays* L.) that excessively high rates of starter fertilizer applied in-furrow could injure plants and this may have accounted for the reduced stands with the in-furrow application of a pop-up fetilizer. They

Table 2. Using soil additives in sesame in the 2016 and 2017 growing seasons

^aAbbreviations: DAP, days after planting.

^bSesame emergence or stand was estimated visually on a scale of 0 to 100 (0 = no emergence and 100 = complete emergence) c These are 3 different rates of the humic acid, organic matter + Bacillus mixture

Table 3. Use of soil additives in sesame for the 2018 growing season

^aAbbreviation: DAP, days after planting

^bStand counts taken 15 DAP. Sesame emergence or stand was estimated visually on a scale of 0 to 100 (0 = no emergence and 100 = complete emergence)

also found that on the lighter sandy loam and silt soils, growth responses with pop-up fertilizer over N alone was primarily due to the P in the pop-up fertilizer. This was probably because of reduced P availability on the sandy, low organic matter, and light colored soils which are typically cold-natured, especially early in the growing season.

3.1.3 2018.*Azospirillum brasilense*, ionized sodium silicate, gibberellic acid, bifenthrin + *Bacillus amyloliquefaciens* strain D747, pyraclostrobin, 2% N, and bifenthrin alone resulted in greater emergence than the untreated check (Table 3). Using *A. brasilense* as an inoculant can affect important changes in the morphology of the plant root. This can be caused by the bacterial production of plant growth regulating substances such as auxin and gibberellins [16,17]. Bolton et al. [18]
reported that A. brasilense did not reported that *A. brasilense* did not consistently increase vegetative growth, turfgrass color, or quality of hybrid bermudagrass [*Cyndon dactylon* (L.) Pers. x *Cynodon transvaalensis* Burtt Davy] compared with the nontreated check.

3.2 Sesame Yield

3.2.1 2016. Gibberellic acid + 3-indolebutyric acid + cytokinin (as kinetin) and 2% N produced yields 32 to 34% higher than the untreated check while the ionized sodium silicate family treatment resulted in a 34% reduction in yield (Table 2). The treatment of 3-indolebutyric acid (0.85%) + cytokinin, as kinetin (0.15%) without gibberellic acid resulted in a 3% yield reduction over the untreated check. Lemus et al [19] reported that using 22.4 and 44.8 kg ha⁻¹ of N produced significantly greater ryegrass (*Lolium multiforum* Lam.) biomass production than the untreated check or gibberellic acid treatment at 29.2 ml ha-1 They speculated that normal growing condition temperatures in the southern US during ryegrass production may be too mild to observe a gibberellic acid response at the applied rates.

3.2.2 2018. Similar results as in 2016 were seen. The 3-way combination of gibberellic acid + 3-indolebutyric acid (0.045%) + cytokinin as kinetin (0.090%) and 2% N resulted in up to a 117% increase in yield over the untreated check (Table 3). No other differences in yield were noted from the untreated check.

4. CONCLUSION

In these studies, the 3-way combination of gibberellic acid + 3-indolebutyric acid (0.045%) + cytokinin as kinetin (0.090%) (sold in the US as Ascend® SL) and 2 % N (sold in the US as Levesol®) proved to be the most consistent soil additives and resulted in a yield increase in the two years that these studies were harvested. The 3-way combination of gibberellic acid $+$ 3indolebutyric acid + cytokinin as kinetin works three different ways. Gibberellic acid stimulates cell division and elongation in leaves and stem, indolebutyric acid stimulates cell division and elongation in leaves and stem while cytokinin promotes cell division and leaf expansion [20]. Also, cytokinin has been found to help in enhancing plant resistance against plant pathogens [21]. The 2% N product has three modes of action: 1) unlocks nutrients in the soil, 2) enhanced nutrient availability results in increased early season growth, overall plant health, and 3) is mobile in the plant for seasonlong activity [22]. It makes phosphorus, zinc, and other key micronutrients more available to the plant and as a result increases early-season growth, overall plant health, and ultimately yield [22].

Using a starter (pop-up) fertilizer either alone or in combination with a fungicide did not greatly influence yield. Variable yield responses have been seen in corn and other crops as well [10, 23-27]. Pierson et al., [24] found that using a starter (pop-up) fertilizer and/or a fungicide in soybean [*Glycine max* (L.) Merr.] was not profitable if soil-borne diseases or nutrient deficiencies were not present. Grichar [10] reported, in a 2-year corn study, that using a pop-up fertilizer + Zn and pop-up fertilizer + Zn + pyraclostrobin in one year resulted inthe highest numerical yields, although this was not significantly different from the untreated check. In the other year, using pop-up fertilizer alone at 28062 and 46771 ml ha⁻¹ resulted in corn yields that were greater than the untreated check.

Azospirillum brasilense resulted in excellent sesame emergence in 2018; however, no improved emergence with *A. brasilense* was shown in 2016 or 2017. Sesame yields were not improved using *A. brasilense*. *A. brasilense* has been used in Brazil with corn as a seed treatment to improve N use and increase yield [28,29]. *A. brasilense* has resulted in an

increase in corn growth and yield when combined with only half of the optimum rate of fertilizer N [28,29]. A meta-analysis of *Azospirillum* spp. indicated that corn yield increases could be achieved when the bacteria was applied without additional N and only minimal increases when applied with N [30].

McFarland [2] had previously found in several studies across the US, that the use of soil additives did not show a significant benefit to crop yield and quality. Laboratory studies have shown that these products did not improve the activity or number of soil microbes and therefore would not be expected to increase the rate or extent of crop residue decomposition [2]. However, El Sawah et al. [31] reported that various components of guar [*Cyamopsis tetragonoloba* (L.)] plant growth including shoot length, root length, leaf area, plant dry weight, nutrient uptake, and yield were significantly affected by the application of biofertilizers and their combination. Soil enzymes activites such as dehydrogenase, phosphatase, protease, and invertase also improved in the soil rhizosphere of plants treated with biofertilizers. They also found that increasing soil enzymes in the rhizosphere and the essential nutrients available for the guar plants increased seed quality by improving the proteins, carbohydrates, starch, fatty acids, and guaran content and reduced the use of chemical fertilizers by 25%.

More work is needed to study the response of crop growth and yield when using soil additives since many new products are constantly being introduced into the market place. Maximum the economic yield of any crop is dependent on using only those inputs which will provide a return on investment.

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COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- 1. McFarland ML, Stichler C, Lemon RG. Non-traditional soil additives: Can they improve crop production? Part 1. Tx. Agric. Ext. Service L-5202. 2002;4.
- 2. McFarland ML. Non-traditional soil additives: Can they improve crop

production? Part 2. Tx. Agric. Ext. Service L-5202. 2006;4.

- 3. Bashir MT, Ali S, Ghauri M, Adris A, Harun R. Impact of excessive nitrogen fertilizers on the environment and associated mitigation strategies. Asian J. Microbiol., Biotechnol., and Environ. Sci. 2013;15(2): 213–221.
- 4. Karlen DL. Tilth. In: Hillel D,editor. Encyclopedia of soils in the environment. Elsevier. 2005;168-174. Accessed 2 October 2023. Available:https://doi.org/10.1016/B0-12- 348530-4/00303-9.
- 5. Bell N, Sullivan DM, Brewer LJ. Hart J. Improving garden soils with organic matter. Oregon State Univ. EC 1561. 2003;16. Accessed 20 July 2023. Available:http://eesc.oregonstate.edu.
- 6. Weaver RW, Dunigan EP, Parr JR, Hiltbold AE. Effect of two soil activators on crop yields and activities of soil microorganisms in the Southern United States. Southern Cooperative Series Bulletin No. 189. Joint Regional Publication by the Agricultural Experiment Stations of Texas, Alabama, Florida, Georgia, Kentucky, Louisiana, North Carolina and Oklahoma; 1974.
- 7. Kocarek M, Kodesova R, Sharipov U, Jursik M. Effect of adjuvant on pendimethalin and dimethenamid-P behavior in soil. J. Hazard. Mater. 2018; 354:266-274.
- 8. Wolkowski, RP, Keeling KA, Oplinger ES. Evaluation of three wetting agents as soil additives for improving crop yield and nutrient availability. Agron. J. 1985;77:695- 698.
- 9. Grichar WJ, Dotray PA, Langham DL. Weed control and the use of herbicides in sesame production. In: Soloneski S, Larramendy ML, editors. Herbicides, Theory, and Application. InTech; 2011. Accessed 29 Sept. 2023. Available:https://doi: 10.5772/12945.
- 10. Grichar WJ, Janak TW, McGinty JA, Brewer MJ. Using biostimulants, soil additives, and plant protectants to improve corn yield in south Texas. Agronomy. 2023;13:1429. Accessed 20 October 2023. Available:https://doi.org/10.3390/agronomy 13051429.
- 11. SAS Institute Incorporated. SAS® Enterprise Guide 8.2 User's Guide. Cary, NC, USA; 2019.

12. Grichar WJ, Woodward JE. Fungicides and application timing for control of early leafspot, southern blight, and *Sclerotinia* blight of peanut. Inter. J. Agronomy. 2016; 7. Article ID 1848723. Accessed 12 October 2023. Available:http://dx.doi.org/10.1155/2016/18

48723. 13. Phipps PM. The response of CBRsusceptible and -resistant cultivars to preplant treatment with metam and infurrow application of Folicur and Abound

for disease management, 2002. F&N

- Reports. 2003;58:FC020. 14. Jordan DL, Brandeburg RL, Bailey JE, Johnson PD, Royals BM, Curtis VL. Compatibility of in-furrow application of acephate, inoculant, and tebuconazole in peanut (*Arachis hypogaea* L.). Peanut Sci. 2006;33:112-117.
- 15. Mascagni HJ, Boquet D, Bell B. Influence of starter fertilizer on corn yield and plant development on Mississippi River alluvial soils. Better Crops; 2007. Accessed 9 October 2023 Available:Ipni.net/publication/bettercrops.n sf/0/1A1444F06D99FESC852579800081D 5BE/\$FILE/Better%20Crops%202007- 2%20p8.
- 16. Hernaandex-Esquivel AA, Castro-Mercado E, Valencia-Cantera E, Alexandre G, Garcia-Pineda E. Application of *Azospirillum brasilense* lipopolysaccharides to promote early wheat plant growth and analysis of related biochemical responses. Front. Sustain. Food Syst. 2020;4:579976. DOI:10.3389/fsufs.2020.579976.
- 17. Fibach-Paldi S, Burdman S, Okon Y. Key physiological properties contributing to rhizosphere adaptation and plant growth promotion abilities of *Azospirillum brasilense*. FEMS Microbiol.lett. 2012;326: 99-108.

DOI:10.1111/j.1574-6968.2011.02407.x

- 18. Bolton C, Cabrera ML, Habteselassie M, Poston D, Henry GM. The impact of commercially available microbial inoculants on bermudagrass establishment, aesthetics, and function. Crop, Forage, & Turfgrass Management. 2022;8(2). Accessed 10 October 2023. Available:https://doi.org/10.1002/cft2.2019 0.
- 19. Lemus R, White JA, and Morrison JI. Effect of gibberellic acid and nitrogen application on biomass and nutritive value of annual

ryegrass. Crop, Forage, & Turfgrass Manage. 2020;7(1). Accessed 20 August 2023.

Available:https://doi.org/10.1002/cft2.2019 Ω .

20. Anonymous. Ascend® SL. Winfield United. 2017;1.

DOI:winfieldunitedag.com

21. Akhtat SS, Pandey C, Mekureyaw MF, Roitsch T. Role of cytokinins for interactions of plants with microbial pathogens and pest insects. Front. Plant Sci. 2019;10. Accessed 10 September 2023. Available:https://doi.org/10.3389/fpls.2019.

01777

- 22. Anonymous. Levesol®: Features and Benefits. CHS Inc. 5500 Cenex Drive. Inver Grove Heights, Mn 55077. 2021;1.
- 23. Niehaus BJ, Lamond RE, Godsey CB, Olsen CJ. Starter nitrogen fertilizer management for continuous no-till corn production. Agron. J. 2004;96:1412-1418. Accessed 26 October 2023. Available:https://doi.org/10.2134/agron/200 4.1412.
- 24. Pierson WL, Kandel YR, Allen TW, Faske TR, Tenuta AU, Wise KA, Mueller DS. Soybean yield response to in-furrow fungicides, fertilizers, and their combination. Crop, Forage & Turfgrass Manage. 2018;4:1-9. Accessed 23 October 2023.

Available:https://doi.org/10.2134/cftm2017. 10.0073

- 25. Rehm GW, Lamb JA. Corn response to fluid fertilizers placed near the seed at planting. Soil Sci. Soc. America J. 2009; 73:1427-1434. Accessed 20 September 2023. Available:https:/doi.org/10.2136/sssaj2008. 0147.
- 26. Wortmann CS, Xerinda SA, Mamo M, Shapiro CA. No-till row crop response to starter fertilizer in eastern Nebraska: I.

Irrigated and rainfed corn. Agron. J. 2006; 98:156-162. Accessed 20 October 2023. Available:https:/doi.org/10.2134/agron/200 5.0015.

27. Wortmann CS, Xerinda SA, Mamo M, Shapiro CA. No-till row crop response to starter fertilizer in eastern Nebraska:II. Irrigated and rainfed corn. Agron. J. 2006;98:187-193. Accessed 26 October 2023.

Available:https:/doi.org/10.2134/agron/200 5.0016.

- 28. Galindo FS, Teixeira Filho MCM, Buzetti S, Pagliari PH, Santini JMK, Alves CJ, Megda MM, Nogueira TAR, Andreotti M, Arf O. Maize yield response to nitrogen rates and sources associated with *Azospirillum brasilense*. Agron. J. 2019; 111:1985-1997.
- 29. Vicente Alves M, Nunes Nest C, Naibo G, Henrique Barreta M, Lazzari M, Florese Junior A, Skoronski E. Corn seed inoculation with *Azospirillum brasilense* in different nitrogen fertilization management. Brazilian J. Agric. Sci. Bras. Ciencias Agrarias*.* 2020;15:1-6.
- 30. Zeffa DM, Fantin LH, dos Santos OJAP, de Oliveira ALM, Canteri MG, Scapim, CA, Goncalves LSA. The influence of topdressing nitrogen on Azospirillum spp. Inoculation in maize crops through meta-analysis. Bragantia. 2018;77:493- 500.
- 31. El-Sawah AM, El-Keblawy A, Ali DFI, Ibrahim HM, El-Sheikh MA, Sharma A, Hamoud YA, Shaghaleh, H, Brestic M, Skalicky, M, Xiong, YC, Sheteiwy MS. Arbuscular mycorrhizal fungi and plant growth-Promoting rhizobacteria enhance soil key enzymes, plant growth, seed yield, and qualitative attributes of guar. Agriculture. 2021;11:194. Accessed 12 October 2023.

Available:https://doi.org/10.3390/agricultur e11030194

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