



Cold Shock Response in Biological Traits of the Silkworm, *Bombyx mori*

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

In response to the ambient temperature, insects' physiology and behaviour change just like those of all other living things. For this investigation, the FC1 X FC2 double hybrid silkworm strain/breed was used. The larvae were collected on the third day of their fifth instar and put in little paper trays for a one-hour cold shock treatment at 10, 15 and 20°C with a relative humidity of 75±5% in B. O. D. The CS-induced larvae were allowed to recuperate for an hour at room temperature. Compared to the control (2.16 g) of the FC1 X FC2 larvae, the average weight of the recorded larvae was 2.30, 2.31, and 2.2 g, which correspond to 10, 15, and 20°C, respectively. When compared to the control group, the population created from 10°C demonstrated an ERR improvement of 70.83 percent (67.50 percent). However, at 15°C and 20°C, respectively, an elevated ERR of 86.67 and 75.83 percent was noted. Mortality was 11.67% between 10 and 15°C, whereas it was 12.50% at 20°C.

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The ability to transition into the next instar, spin a cocoon, become a pupa and eventually become a moth was utilised to assess the cold's potential impact on larval, pupal, and adult mortality. Highest cocoon weight of 1.37 g was observed at 15°C. Weights of the cocoons were measured and compared to the control (1.30 g), which corresponds to temperatures of 10 and 20°C, respectively. The cocoons made by CS larvae at 10, 15, and 20°C and the control had shell weights of 0.22, 0.24, and 0.19 g, compared to 0.30 g for the control. Temperature of 15°C showed higher efficiency in all the traits studied whereas other temperatures showed a slight decline in all the traits. As a result, we inferred that FC1 X FC2 had exhibited a profound response to CS temperature of 15°C and can be used to develop CS silkworm strains for the temperate areas.

Keywords: Cold; shock; biological; commercial; traits; *Bombyx mori*.

1. INTRODUCTION

Silkworms are one of the most significant domesticated insects, producing luxurious silk thread in the form of cocoons by eating mulberry leaves during their larval stage. Environmental factors have a significant impact on silkworm growth and development. Ambient temperature, rearing seasons, quality mulberry leaf and silkworm strain genetic makeup all influence on biological and cocoon-related characteristics. Temperature fluctuation during different phases of larval development, on the other hand, was found to be better for larvae growth and development than constant temperature. Many studies have shown that good quality cocoons are created at temperatures between 22 and 27°C and that temperature above these values result in low-grade cocoons [1]. Chill-sensitive insects die as a result of cold-induced damage before ice forms within their bodies. Cold-intolerance is a term used to describe this method. False codling moth larvae, *Thaumatotobia leucotreta* (Lepidoptera: Tortricidae), for example, freeze between 13 and 22°C but are killed by brief exposures between 8 and 12°C. [2] discovered that lower temperatures are always better than higher temperatures. Cold shock is the stress caused by a quick and rapid exposure to cold temperatures that are not below freezing. Cold shock, also known as "direct chilling injury," is caused by a rapid cooling rate. The temperature threshold that causes injury varies by species and strain, but in the absence of ice formation and at temperatures considerably over the super cooling limit, this type of injury is regularly found. Insects are continually confronted with adverse environmental conditions such as pathogen infections, UV radiation, pesticide activity, oxidative stress and extreme temperature. Insects, on the other hand, can cold-harden in a significantly shorter time frame, a process known

as fast cold-hardening [3]. As a result of these procedures, the influence of cold shock on larval biology and commercial features is unclear. The goal of the coordinated efforts to improve the cocoon characteristics of domesticated silkworms was to produce superior quality silk. As a result, developing bivoltine breeds/hybrids that can tolerate low temperature stress conditions, particularly in temperate zones, becomes crucial. So, the current study was designed to see how low temperatures affected several biological features in bivoltine hybrids of *B. mori*.

2. MATERIALS AND METHODS

The FC1 X FC2 double hybrid silkworm strain/breed was chosen for this study. The Department of Sericulture, Jammu and Kashmir Union Territory, provided first instar (first moulting) larvae. In the laboratory, the first instar (first moulting) larvae were incubated at a temperature of 25±1°C and a relative humidity of 75±5%. The larvae were reared on mulberry leaves until they spun cocoons, as per conventional protocol [4]. The larvae were collected during the 5th instar (3rd day) and placed in small paper trays for cold shock treatment at 10, 15 and 20°C with a relative humidity of 75% in B. O. D for 1 hour. The CS-induced larvae were kept at room temperature for 1 hour to recover.

2.1 Analysis of Biological and Commercial Traits

2.1.1 Determination of cold sensitivity

The capacity to enter the next instar or spin cocoon, transform into pupae and moth was used to determine cold sensitivity in terms of larval, pupal and adult mortality.

2.1.2 Effective Rate of Rearing (ERR)

The following formula was used to determine ERR,

$$\text{ERR} = \frac{\text{Number of good cocoons spun}}{\text{Number of larvae brushed}} \times 100$$

2.1.3 Larval, cocoon, pupal and shell weight

On day 6 of the fifth instar, approximately 6 larvae were randomly selected from each replication and their weight was recorded. Similarly, from each replication on day 6 after spinning, cocoon weight was determined. The pupae retrieved from randomly selected cocoons (6) of each replication were used to calculate pupal weight and after removing pupae, shells were weighed for cocoon shell weight.

2.1.4 Shell ratio

The following formula was used to calculate the shell ratio,

$$\text{Shell ratio} = \frac{\text{Shell weight}}{\text{Cocoon weight}} \times 100$$

2.1.5 Emergence of moth and fecundity

All of the cocoons derived from the various CS and control batches were kept in normal environmental conditions until eclosion. The pupal and adult mortality rates as impacted by the CS during different treatments were calculated using the moth that had emerged. The average number of eggs laid by 6 moths from each treatment was used to calculate fecundity. The eggs were counted and documented separately.

The data generated was put to statistical tool, one way ANOVA for analysis and interpretation of data by using SPSS (Statistical Package Software for Social Sciences) version 20.

3. RESULTS

3.1 Changes in the Larval Growth due to Cold Shock

The weight of the larvae on day 3 of the fifth instar was used to determine how CS affected their growth at different temperatures. As a result, the average weight of the larvae recorded was 2.30, 2.31, and 2.2 g which corresponds to 10, 15 and 20°C, respectively, compared to the control (2.16 g) of the FC1 X FC2 larvae (Table 1), which was statistically significant at $P < 0.01$. At all temperatures, however, there was a modest increase in larval weight. Interestingly, the larvae produced from 15°C CS weighed 2.31 g more than the control larvae (2.31 g) (Fig. 1).

3.2 Changes in the ERR due to Cold Shock

The ERR stands for the larvae that successfully spin cocoons. In the end, silkworm larvae derived from various CS induced were grown in the rearing house under natural environmental circumstances. Surprisingly, the population formed from 10°C showed a 70.83 % improvement in ERR when compared to the control group (67.50 %). However, at 15°C, an increased ERR of 86.67 percent was seen, and at 20°C, an increased ERR of 75.83 % was observed (Table 1, Fig. 2), which is significant at $P < 0.01$.

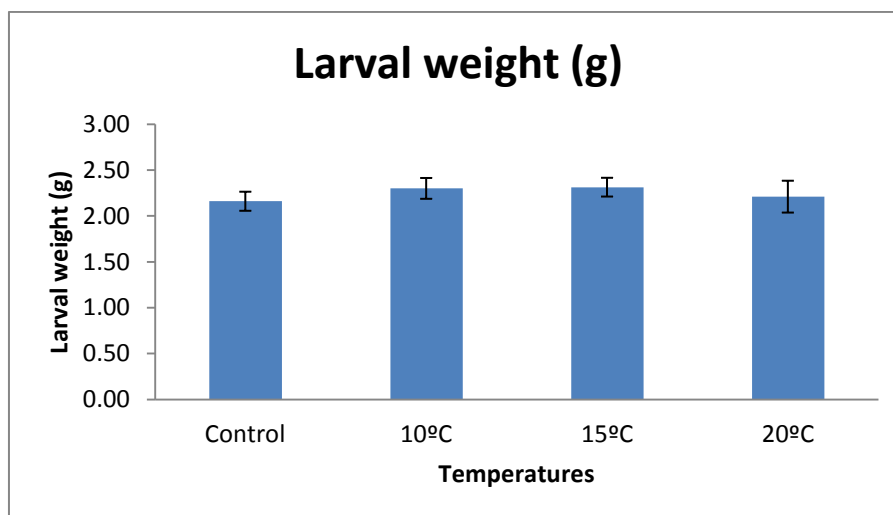


Fig. 1. Larval weight FC1 X FC2 larvae at different temperatures

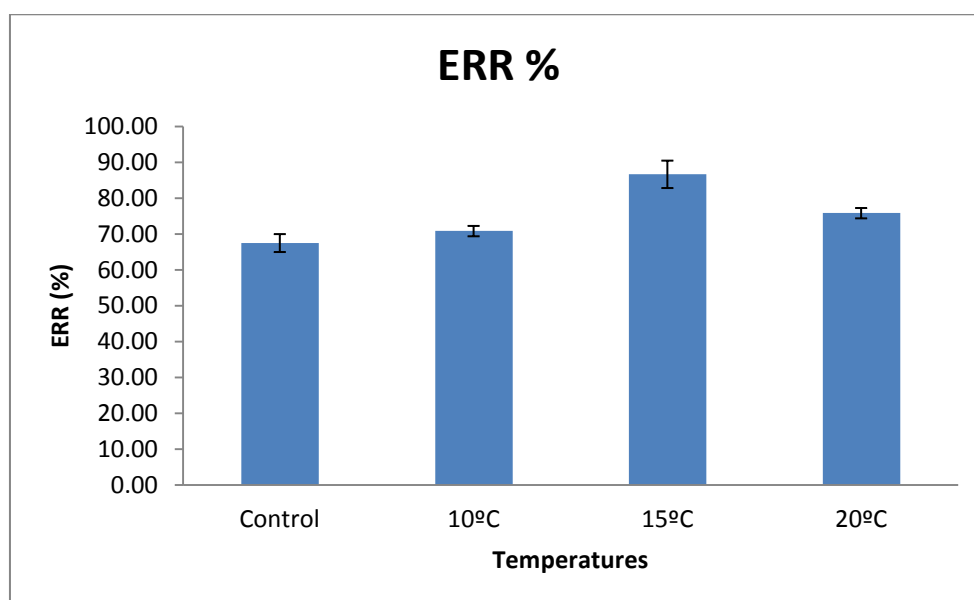


Fig. 2. ERR of FC1 X FC2 larvae at different temperatures

3.3 Changes in the Larval Mortality due to Cold Shock

FC1 X FC2 larvae are extremely sensitive to different cold shock (CS) temperatures (Fig. 3). The FC1 X FC2 larvae were found to be sensitive to the CS temperatures of 10, 15, and 20°C. At 10 and 15°C, mortality was 11.67 percent, but at 20°C, it was 12.50 percent (Table 1), which is significant at $P < 0.01$.

3.4 Changes due to Cold Shock in Relation to Cocoon Characters

3.4.1 Cocoon weight

The weight of the cocoons spun by FC1X FC2 silkworm larvae produced from CS induced on day 6 at 10, 15 and 20°C was significantly higher than the control (Table 2, Fig. 4). At 15°C, the cocoon weighed more (1.37 g). In comparison to the control (1.30 g), cocoon weights of 1.29 and 1.23 g were measured, which correspond to 10 and 20°C, respectively (Table 2).

3.4.2 Shell weight

Due to the fluctuating environmental conditions in the rearing house, the cocoon shell weight was clearly altered, just like the cocoon weight in control. As a result, the weight of the cocoon shell in the control group was 0.33 g. However, cold shock induced larvae at 15°C showed a considerable improvement in shell weight. The

shell weight of the cocoons generated by CS larvae at 10, 15, 20°C and control was 0.22, 0.24, and 0.19 g respectively (Table 2, Fig. 5), compared to 0.30 g for the control.

3.4.3 Pupal weight

Surprisingly, the weight of the pupa, as a measure of its growth, was highest in the population obtained from CS at 15 and 10°C on day-6. Pupal weight of 1.03 and 0.99 g of pupal weight were observed at 20 °C and control (Table 2).

3.4.4 Cocoon shell ratio

The cocoon shell weight ratio was similarly adjusted as the cocoon and shell weight in fifth instar larvae in FC1 X FC2. The control population had a cocoon shell ratio of 23.73 %, while the populations generated from FC1 X FC2 CS larvae at 10, 15, and 20°C had shell ratios of 17.67, 17.26 and 16.01 %, respectively (Table 2, Fig. 7).

3.4.5 Moth emergence (%)

In the FC1 X FC2 double hybrid silkworm strain, moth emergence was similarly adjusted as other parameters changed. Moth emergence was 89.04 percent in the control population, while moth emergence was 94.16, 96.18, and 95.28 percent in the FC1 X FC2 double hybrid silkworm strain CS populations grown at 10, 15, and 20°C, respectively (Table 3, Fig. 8).

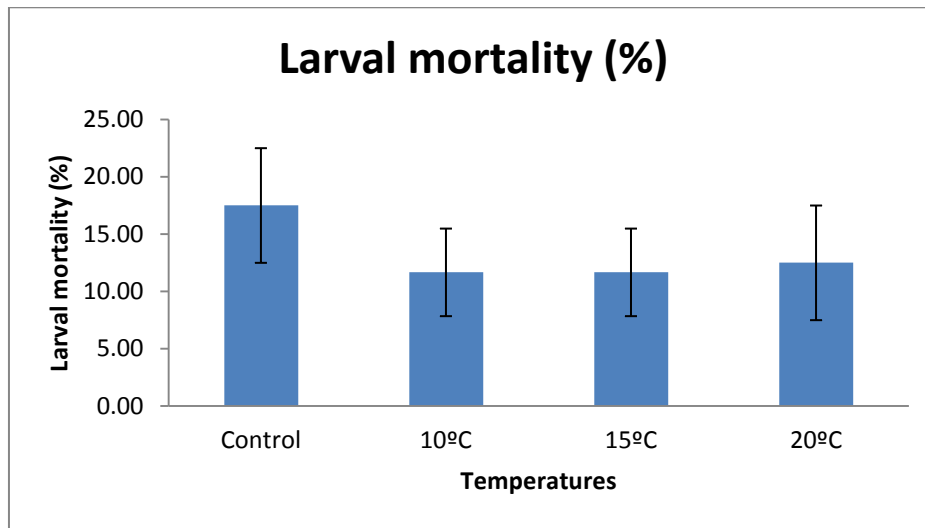


Fig. 3. Larval mortality of FC1X FC2 at different temperatures

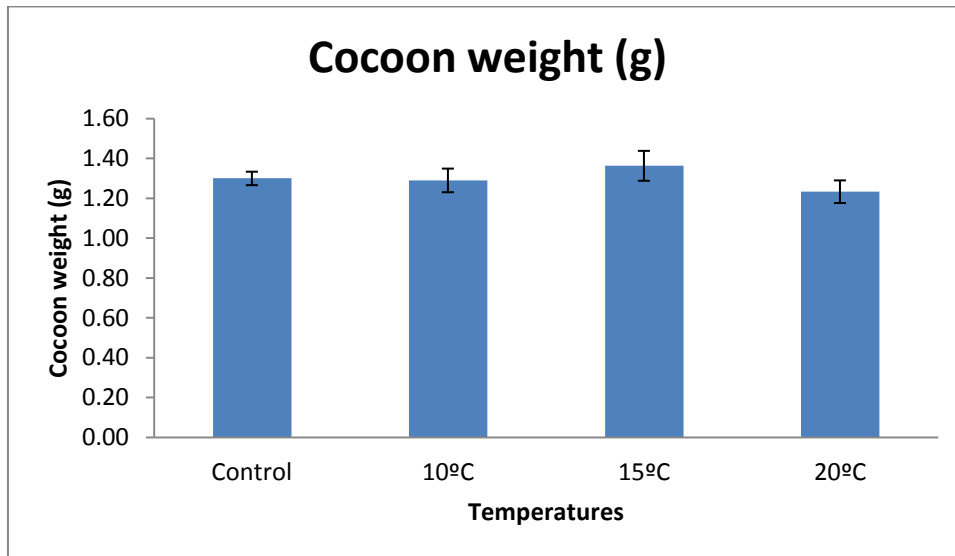


Fig. 4. Cocoon weight of FC1 X FC2 larvae at different temperatures

Table 1. Effect of cold shock on biological traits of the silkworm, *Bombyx mori* bivoltine hybrid FC1 X FC2

Treatments	Larval weight (g) Mean±S.E.	ERR (%) Mean±S.E.	Larval mortality (%) Mean±S.E.
Control	2.160±0.060	67.500±1.443	17.500±2.887
10°C	2.300±0.066	70.833±0.833	11.667±2.205
15°C	2.313±0.059	86.667±2.205	11.667±2.205
20°C	2.210±0.100	75.833±0.833	12.500±2.887
C.D.	N/A	4.780	N/A
SE(m)	0.073	1.443	2.569
SE(d)	0.103	2.041	3.632
C.V.	5.642	3.324	33.366
F-Value	1.005	33.639	1.193
Significance	0.43906	0.00007	0.37228

Table 2. Effect of cold shock on cocoon traits of the silkworm, *Bombyx mori* bivoltine hybrid FC1 X FC2

Treatments	Cocoon weight (g)	Pupal weight (g)	Shell weight (g)	Shell ratio (%)
	Mean±S.E.	Mean±S.E.	Mean±S.E.	Mean±S.E.
Control	1.300±0.019	0.987±0.015	0.303±0.009	23.733±0.333
10°C	1.290±0.034	1.053±0.007	0.260±0.006	17.667±2.539
15°C	1.363±0.043	1.127±0.019	0.330±0.012	17.257±3.792
20°C	1.233±0.033	1.033±0.015	0.193±0.035	16.013±2.582
C.D.	N/A	0.047	0.064	N/A
SE(m)	0.034	0.014	0.019	2.627
SE(d)	0.047	0.020	0.027	3.715
C.V.	4.484	2.349	12.301	24.376
F-Value	2.517	16.731	9.562	1.724
Significance	0.13186	0.00083	0.00505	0.23905

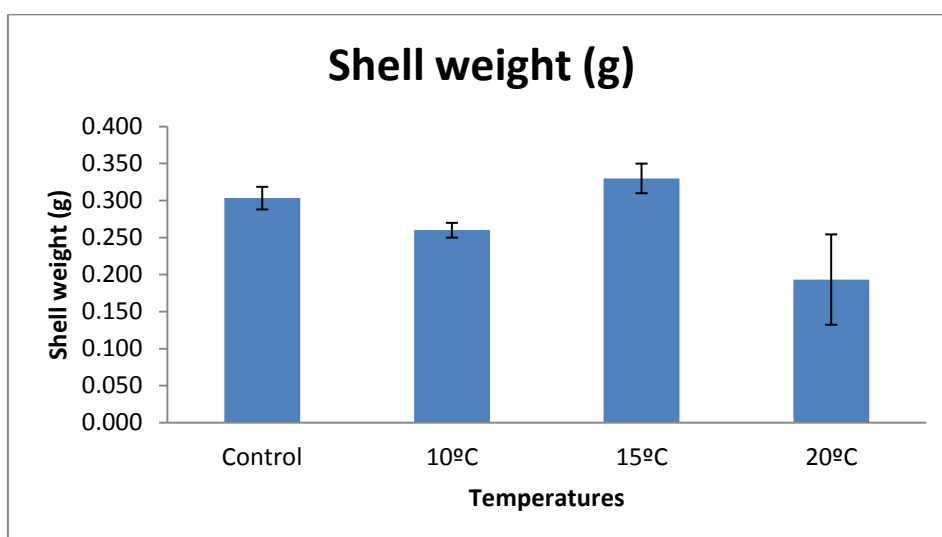


Fig. 5. Cocoon shell weight of FC1 X FC2 larvae at different temperatures

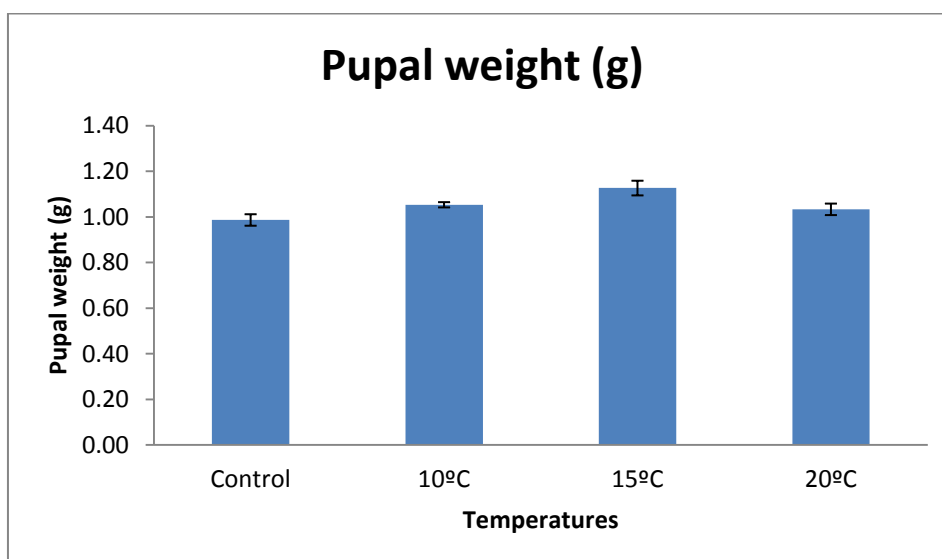


Fig. 6. Pupal weight of FC1 X FC2 larvae at different temperatures

3.4.6 Fecundity (No. of eggs laid)

Fecundity altered in the FC1 X FC2 double hybrid silkworm strain as other parameters changed. The control moth laid 446.00 eggs,

while the FC1 X FC2 double hybrid silkworm strain CS populations reared at 10, 15, and 20°C laid 438.33, 559.00, and 460.00 eggs respectively (Table 3, Fig. 9).

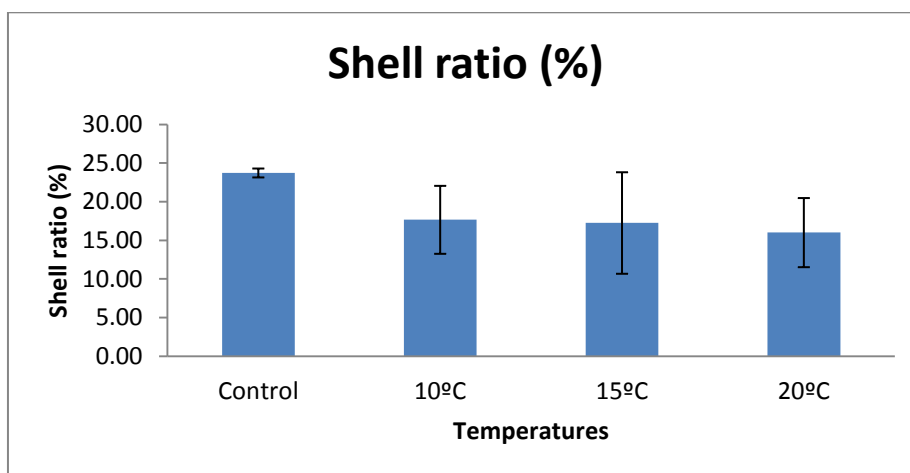


Fig. 7. Shell ratio of FC1 X FC2 larvae at different temperatures

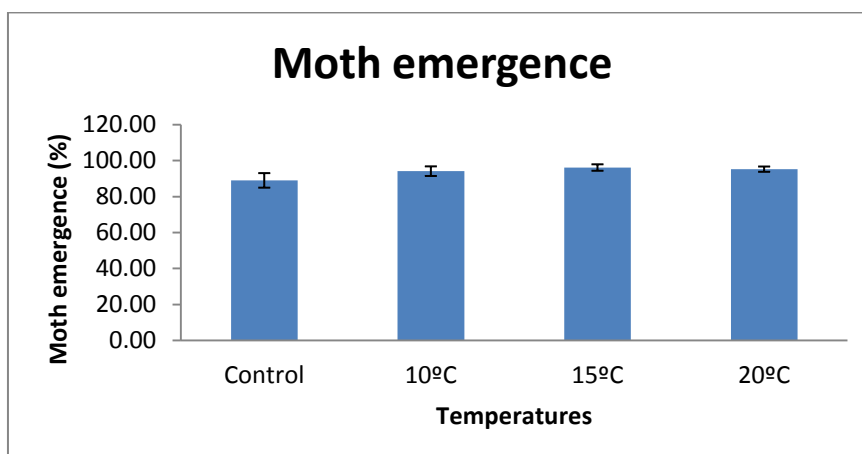


Fig. 8. Moth emergence (%) of FC1 X FC2 double hybrid at different temperatures

Table 3. Effect of cold shock on Moth emergence of the silkworm, *Bombyx mori* bivoltine hybrid FC1 X FC2

Treatments	Moth emergence (%)	Fecundity
	Mean±S.E.	Mean±S.E.
Control	89.040±2.335	446.000±8.718
10°C	94.157±1.545	438.333±10.929
15°C	96.180±1.036	559.000±9.539
20°C	95.277±0.846	460.000±5.774
C.D.	5.139	29.611
SE(m)	1.552	8.941
SE(d)	2.194	12.645
C.V.	2.869	3.255
F-Value	4.230	39.459
Significance	0.04566	0.00004

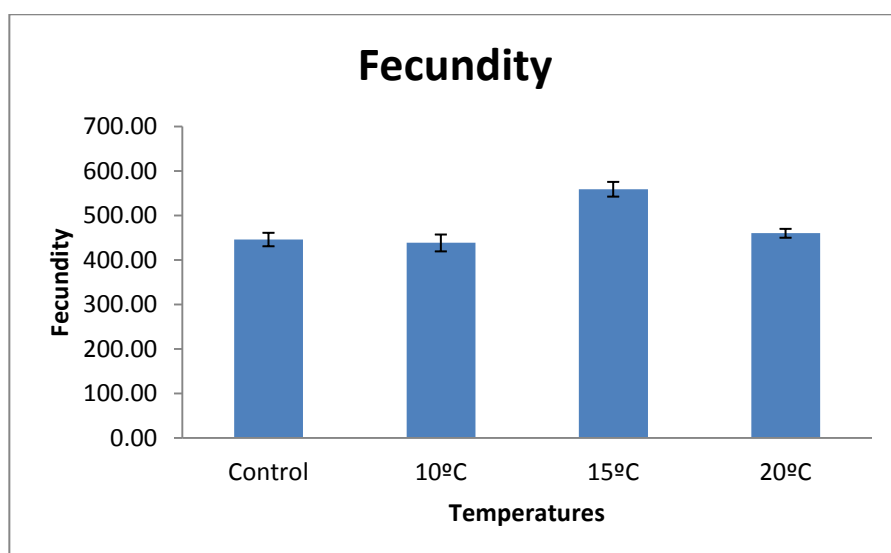


Fig. 9. Number of eggs laid by female moth of FC1 X FC2 double hybrid at different temperatures

4. DISCUSSION

Because the insects were rapidly cooled and spontaneous ice formation in extracellular fluids did not begin until around 15 °C lower, these data indicate that mortality was caused by cold shock [5,3]. The average weight of the larvae observed was 2.30, 2.31 and 2.2 g, respectively, corresponding to 10, 15 and 20°C, as opposed to the control (2.16 g) of the FC1 X FC2 larvae (Table 1), which was statistically significant at $P < 0.01$. The weight of the larvae increased somewhat at all temperatures. Surprisingly, the 15°C CS larvae weighed 2.31 g greater than the control larvae. This response was first described in larvae, pupae, and pharate adults of both diapause and nondiapause meat flies, *Sarcophaga crassipalpis*, by [3]. Non-overwintering stages of a beetle, *Xanthogaleruca luteola*, a true bug, *Oncopeltus fasciatus* and another fly, *Sarcophaga bullata*, exhibit this exceptionally quick cold-hardening reaction [5]. In a study of a tephritid fruit fly, [6] looked at temperature acclimatization in relation to and flying thresholds. He said that *Dacus* try one may acclimate quickly, even at cooling rates. *D. melanogaster* is a cosmopolitan species that can be found in all biogeographic zones [5]. FC1 X FC2 larvae are very sensitive to various cold shock (CS) temperatures. The CS temperatures of 10, 15, and 20°C were found to be responsive to the FC1 X FC2 larvae. Mortality was 11.67 % at 10 and 15°C, but 12.50 % at 20°C, which is significant at $P < 0.01$. *D. melanogaster* is a global species that can be found in all

biogeographic zones [7]. Among the six species of the melanogaster subgroup, this species is said to be the most resistant to the environmental pressures of high-temperature desiccation and low temperature [8]. The most common fruit fly, *D. melanogaster*, is commonly employed as an experimental organism for genetics research; nevertheless, it is unknown how this organism overwinters in temperate climates [7]. The cocoon weight spun by FC1 X FC2 silkworm larvae induced on day 6 at 10, 15, and 20°C was significantly higher than the control. The cocoon weighed was highest at 15°C (1.37 g). The cocoon shell weight, like the cocoon weight in control, was clearly affected by changing environmental conditions in the rearing house. The weight of the cocoon shell in the control group was 0.33 g as a result. Cold shock induced larvae at 15°C, on the other hand, showed a significant improvement in shell weight. The shell weight of the cocoons produced by CS larvae at 10, 15, 20°C and control was 0.22, 0.24 and 0.19 g respectively, compared to 0.30 g for the control. The pupal weight as a metric of growth was highest in the population derived from CS on day-6 at 15 and 10°C. At 20°C and control, pupal weights of 1.03 and 0.99 g were reported during the 6th day larval stage of FC1 X FC2. The cocoon shell weight ratio was similarly adjusted as the cocoon and shell weight in fifth instar larvae in FC1 X FC2. The control population had a cocoon shell ratio of 23.73%, while the populations generated from FC1 X FC2 CS larvae at 10, 15, and 20°C had shell ratios of 17.67, 17.26, and 16.01 %, respectively. The

FC1 X FC2 double hybrid silkworm strain's fecundity fluctuated as other factors changed. The FC1 X FC2 double hybrid silkworm strain CS populations raised at 10, 15 and 20°C deposited 438.33, 559.00, and 460.00 eggs, respectively. The average weight of the larvae observed was 2.30, 2.31 and 2.2 g, respectively, corresponding to 10, 15 and 20°C, as opposed to the control (2.16 g) of the FC1 X FC2 larvae (Table 1), which was statistically significant at $P < 0.01$. The weight of the larvae increased somewhat at all temperatures. Surprisingly, the 15°C CS larvae weighed 2.31 g greater than the control larvae. As a form of defence against cold shock, insects appear to have a high potential for fast cold hardening. Understanding the underlying concepts of insect cold tolerance and the nature of cold injury is required for properly understanding the physiological mechanisms driving cold-hardening. The super cooling point (SCP) is a measure of the lower lethal temperature in some insects, however significant mortality in *D. melanogaster* at temperatures considerably the SCP was observed. The SCPs of *D. melanogaster* larvae, pupae, and adults were in the -17 to -20°C range. Previous investigations of cold tolerance in *drosophilid* species [9,10] have found similar results.

5. CONCLUSION

The recent decade has seen a huge rise in studies addressing the processes of cold hardening thanks to developments in molecular biology and the 'omics' revolution. Rapid cold hardening (RCH) normally occurs at temperatures below the developmental threshold of most species, whereas cold acclimation occurs at temperatures that allow for growth and reproduction. In contrast to cryoprotectant mobilisation, both seasonal cold-hardening and RCH are thought to include cell membrane changes. The function of gene expression is perhaps the most fundamental mechanistic difference between seasonal cold-hardening and RCH. Furthermore, further functional research is needed to determine which processes and pathways are required for cold hardening and which just correlate with increased cold tolerance. Moth emergence was also altered as other parameters changed in the FC1 X FC2 double hybrid silkworm strain. The control population had an emergence rate of 89.04 %, while the FC1 X FC2 double hybrid silkworm strain CS populations reared at 10, 15, and 20°C had emergence rates of 94.16, 96.18, and 95.28 %, respectively. As other variables changed, the

fertility of the FC1 X FC2 double hybrid silkworm strain fluctuated. The CS populations of the FC1 X FC2 double hybrid silkworm strain laid 438.33, 559.00, and 460.00 eggs, respectively, when grown at 10, 15, and 20°C.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Suresh Kumar N, Harjeet Singh. Expression of heterosis in silkworm hybrids, *Bombyx mori* (*Lepidoptera: Bombycidae*) tolerant to high temperature and high and low humidity conditions of the tropics. International Journal of Plant, Animal and Environmental Sciences. 2011;1:3;188-204.
2. Datta RK, Suresh Kumar N, Basavaraja HK, Kishor Kumar CM., Mal Reddy N. On the breeding of "CSR18 × CSR19" A robust bivoltine hybrid of Silkworm, *Bombyx mori*, for the tropics. International Journal of Industrial Entomology. 2002;5:2: 153-162.
3. Lee RE, Chen CP, Denlinger DL. A rapid cold-hardening process in insects. Science. 1987;238:1415-1417.
4. Jolly MS. Appropriate sericulture techniques. Gitanjali Printers. 1987;63-106.
5. Chen C, Walker V. Increase in cold shock tolerance by selection of cold resistant lines in *Drosophila melanogaster*. Ecological Entomology. 2008;18;3:184-190.
6. Meats A. Rapid acclimatization to low temperature in the Queensland fruit fly *Dacus tryoni*. Journal of Insect Physiology. 1973;19:1903-1911.
7. David JR, Allemand R, vanherrewege J, Cohet Y. Ecophysiology: Abiotic factors in the genetics and biology of *Drosophila*. New York: Academic Press. 1983;3;105-170.
8. Stanley SM, Parsons PA, Spence GE, Weber L. Resistance of species of the

- Drosophila melanogaster* subgroup to environmental extremes. Australian Journal Zoology. 1980;28:413-421.
9. Tucic N. Genetic capacity for adaptation to cold resistance at different developmental stages of *Drosophila melanogaster*. Evolution. 1979;33:350-358.
10. Enomoto O. Larval diapause in *Chymomyza costata* (Diptera: *Drosophilidae*). II. Frost avoidance. Low Temperature. Science. 1981;39:31-39.

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